

Application	Amount	Volume (μL)	Conc. (ng/ μL)	Quality	Size (bp)	Comment
Illumina Short-Read Sequencing						
Genome Sequencing						
<i>Whole Genome - Low input (incl. PCR)</i>	50 – 1000 ng	10 – 50	>2	high molecular for best results 260/280 ratio 1.8-2.0	-	FFPE possible with constraints in output quality
<i>Whole Genome - PCR Free</i>	1.25 – 2.5 μg	10 – 50	>25	high molecular for best results 260/280 ratio 1.8-2.0	-	standard is 350 bp insert size, for 550 bp insert size >2000 ng
<i>Whole Exome Sequencing</i>	> 50 ng	10	5	260/280 ratio 1.8-2.0	-	FFPE possible if amount based on qPCR measurement is sufficient
<i>Gene Panel</i>	600 ng	10	20	260/280 ratio 1.7-2.2 260/230 ratio 1.2	-	-
<i>Amplicons</i>	> 50 ng	10	5	260/280 ratio >1.5	-	less material possible on request
Microbiome						
<i>Microbiome 16S</i>	100 ng	10	10	260/280 ratio >1.5	-	less material possible on request
<i>Microbiome Shotgun Metagenomics</i>	50 ng	10	5	260/280 ratio >1.5	-	less material possible on request

Application	Amount	Volume (μ L)	Conc. (ng/ μ L)	Quality	Size (bp)	Comment
Illumina Short-Read Sequencing						
<i>Transcriptome</i>						
<i>Whole Transcriptome – standard input</i>	500 ng	10	50	260/280 ratio >2.0 RQN \geq 8; Δ <1 DNase treated	-	-
<i>Whole Transcriptome – low input</i>	1 ng	10	<1	260/280 ratio >2.0 RQN \geq 8; Δ <1 DNase treated	-	constraints in output quality possible if input amount is very low
<i>Expression Profiling (mRNA) – standard input</i>	500 ng	10	50	260/280 ratio >2.0 RQN \geq 8; Δ <1 DNase treated	-	-
<i>Expression Profiling (mRNA) – low input</i>	1 ng	10	<1	260/280 ratio >2.0 RQN \geq 8; Δ <1 DNase treated	-	constraints in output quality possible if input amount is very low
<i>Small RNA Profiling (miRNA, lnc-RNA etc.)</i>	50 ng	10	50	260/280 ratio >2.0 RQN \geq 8; Δ <1 DNase treated	-	-
<i>Sequencing of prepared NGS libraries</i>	50 ng	10	5	No primer dimer residuals	-	Buffer: TRIS-HCL 10 mM

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DNA-Microarray						
<i>Transcriptome Profiling – standard input</i>	500 ng	10	50	260/280 ratio 1.8 – 2.0 260/230 ratio >2.0 DNase treated RQN \geq 8; Δ <1	-	-
<i>Transcriptome Profiling – low input</i>	>0.5 ng	10	>0.05	260/280 ratio 1.8 – 2.0 260/230 ratio >2.0 DNase treated RQN \geq 8; Δ <1	-	constraints in output quality possible if input amount is very low
10X Single-Cell RNA-Sequencing						
<i>Single-Cell Transcriptomics</i>	1.000 – 30.000 cells /sample	50 μl	500-1.000 cells/ μl	Vitality > 70 %	-	-
<i>Spatial Transcriptomics</i>	Tissue sections placed on Visium slides, RNA extracted from tissue section RIN > 7					
Bionano Optical Mapping						
<i>Bionano Saphyr Chip</i>	1 μg	-	> 36	260/280 = 1.8 260/230 = 2.0-2.2	Megabase range	-

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Pacific Biosciences Long-Read Sequencing						
<i>Transcriptome (Iso-Seq)</i>	2 μg	20	100	RIN 8-10 260/280 = 2.0 260/230 = 2.2	-	DNase digested
<i>Whole genome sequencing</i>						
<i>HiFi reads – standard protocol</i>	15 μg	-	-	260/280 = 1.8 260/230 = 2.0-2.2	>50kb	HMW DNA, RNase digested
<i>HiFi reads – Low Input</i>	1 μg	-	-	260/280 = 1.8 260/230 = 2.0-2.2	>50kb	HMW DNA, RNase digested
<i>HiFi reads – Ultra-Low Input</i>	20 ng	-	-	-	>50kb	HMW DNA, RNase digested
<i>CLR reads > 30 kb</i>	8 μg	-	-	260/280 = 1.8 260/230 = 2.0-2.2	>50kb	HMW DNA
<i>CLR reads > 15 kb</i>	5 μg	-	-	260/280 = 1.8 260/230 = 2.0-2.2	>50kb	HMW DNA
<i>CLR reads ~ 10 kb</i>	1.5 μg	-	-	260/280 = 1.8 260/230 = 2.0-2.2	>20kb	-
<i>Amplicon sequencing</i>	500 ng – 3 μg (depends on size)	50	-	-	-	Clean, target-specific
<i>Multiplexed Microbial</i>	1 μg per sample	20-100	-	260/280 = 1.8 260/230 = 2.0-2.2	>20kb	-

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Oxford Nanopore Long-Read Sequencing

<i>Transcriptome</i>						
<i>Direct mRNA Sequencing</i>	>500 ng polyA+	> 10	> 60	260/280 = 2.0 260/230 = 2.0-2.2	-	-
<i>cDNA Sequencing</i>	>100 ng polyA+	> 10	> 15	260/280 = 2.0 260/230 = 2.0-2.2	-	200 ng of already prepped cDNA can also be used as input
<i>cDNA PCR Sequencing</i>	>1 ng polyA+ or > 50 ng Total-RNA	> 10	> 1	260/280 = 2.0 260/230 = 2.0-2.2	-	-
<i>Whole Genome Sequencing</i>						
<i>Ligation Sequencing</i>	> 2 μg	10-50	> 40	260/280 = 1.8 260/230 = 2.0-2.2	> 30 kb	-
<i>Native Barcoding</i>	> 1 μg per sample	10-50	> 40	260/280 = 1.8 260/230 = 2.0-2.2	> 30 kb	-
<i>Ultra Long Reads</i>	20 μg	20	1000	260/280 = 1.8 260/230 = 2.0-2.2	-	-
<i>Genome Rapid</i>	> 400 ng	10	> 55	260/280 = 1.8 260/230 = 2.0-2.2	-	-
<i>PCR Sequencing</i>	> 100 ng	> 10 - 50	> 2	260/280 = 1.8 260/230 = 2.0-2.2	-	-

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Oxford Nanopore Long-Read Sequencing

<i>Amplicon Sequencing</i>						
<i>Amplicons by Ligation</i>	> 1 μg per sample	10-50	> 40	260/280 = 1.8 260/230 = 2.0-2.2	-	-
16S	> 10 ng	> 10	> 1	260/280 = 1.8 260/230 = 2.0-2.2	-	-