

Application	Amount	Volume (μL)	Conc. ($\text{ng}/\mu\text{L}$)	Quality	Size (bp)	Comment
Illumina Short-Read Sequencing						
Genome Sequencing						
<i>Whole Genome - Low input (incl. PCR)</i>	50 – 1000 ng	10 – 50	>2	high molecular for best results 260/280 ratio 1.8-2.0	-	FFPE possible with constraints in output quality
<i>Whole Genome - PCR Free</i>	1.25 – 2.5 μg	10 – 50	>25	high molecular for best results 260/280 ratio 1.8-2.0	-	standard is 350 bp insert size, for 550 bp insert size >2000 ng
<i>Whole Exome Sequencing</i>	>50 ng	10	5	260/280 ratio 1.8-2.0	-	FFPE possible if amount based on qPCR measurement is sufficient
<i>Gene Panel</i>	600 ng	10	20	260/280 ratio 1.7-2.2 260/230 ratio 1.2	-	-
<i>Amplicons</i>	>50 ng	10	5	260/280 ratio >1.5	-	less material possible on request
Microbiome						
<i>Microbiome 16S</i>	100 ng	10	10	260/280 ratio >1.5	-	less material possible on request
<i>Microbiome Shotgun Metagenomics</i>	50 ng	10	5	260/280 ratio >1.5	-	less material possible on request

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Illumina Short-Read Sequencing						
<i>Transcriptome</i>						
<i>Whole Transcriptome – standard input</i>	500 ng	10	50	260/280 ratio >2.0 RQN \geq 8; Δ <1 DNase treated	-	-
<i>Whole Transcriptome – low input</i>	1 ng	10	<1	260/280 ratio >2.0 RQN \geq 8; Δ <1 DNase treated	-	constraints in output quality possible if input amount is very low
<i>Expression Profiling (mRNA) – standard input</i>	500 ng	10	50	260/280 ratio >2.0 RQN \geq 8; Δ <1 DNase treated	-	-
<i>Expression Profiling (mRNA) – low input</i>	1 ng	10	<1	260/280 ratio >2.0 RQN \geq 8; Δ <1 DNase treated	-	constraints in output quality possible if input amount is very low
<i>Small RNA Profiling (miRNA, lnc-RNA etc.)</i>	50 ng	10	50	260/280 ratio >2.0 RQN \geq 8; Δ <1 DNase treated	-	-
<i>Sequencing of prepared NGS libraries</i>	50 ng	10	5	No primer dimer residuals	-	Buffer: TRIS-HCL 10 mM

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Gene Expression Microarray						
<i>Transcriptome Profiling – standard input</i>	500 ng	10	50	260/280 ratio 1.8 – 2.0 260/230 ratio >2.0 DNase treated RQN \geq 8; Δ <1	-	-
<i>Transcriptome Profiling – low input</i>	>0.5 ng	10	>0.05	260/280 ratio 1.8 – 2.0 260/230 ratio >2.0 DNase treated RQN \geq 8; Δ <1	-	constraints in output quality possible if input amount is very low
10X Single-Cell RNA-Sequencing						
<i>Single-Cell Transcriptomics</i>	1.000 – 30.000 cells /sample	50 μl	500-1.000 cells/ μl	Vitality >70 %	-	-
<i>Spatial Transcriptomics</i>	Tissue sections placed on Visium slides, RNA extracted from tissue section RIN >7					
Bionano Optical Mapping						
<i>Bionano Saphyr Chip</i>	1 μg	-	>36	260/280 = 1.8 260/230 = 2.0-2.2	Megabase range	-

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Pacific Biosciences Long-Read Sequencing						
<i>Transcriptome (Iso-Seq)</i>	>2 μg	>15	>130	RIN 8-10 260/280 = 2.0 260/230 = 2.2	-	DNase digested, buffer: RNase free water
<i>Whole Genome Sequencing</i>						
<i>HiFi Reads – Standard Protocol</i>	>6 μg	>120	50	260/280 = 1.8 260/230 = 2.0-2.2	>50kb	homogenous HMW DNA, RNase digested, buffer: EB
<i>HiFi Reads – Low Input</i>	>1 μg	>50	20	260/280 = 1.8 260/230 = 2.0-2.2	>50kb	homogenous HMW DNA, RNase digested, buffer: EB
<i>HiFi Reads – Ultra-Low Input</i>	>30 ng	>15	2	-	>50kb	homogenous HMW DNA, RNase digested, buffer: EB
<i>Continous Long Reads</i>	>6 μg	>120	50	260/280 = 1.8 260/230 = 2.0-2.2	>50kb	homogenous HMW DNA, RNase digested, buffer: EB
<i>Amplicon Sequencing</i>	500 ng – 3 μg (depends on size)	50	-	-	-	clean, target-specific, buffer: EB
<i>Multiplexed Microbial</i>	>1 μg per sample	20-100	-	260/280 = 1.8 260/230 = 2.0-2.2	>20kb	-

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Oxford Nanopore Long-Read Sequencing						
<i>Transcriptome</i>						
<i>Direct mRNA Sequencing</i>	>600 ng polyA+	>10	>60	260/280 = 2.0 260/230 = 2.0-2.2	-	DNase digested, buffer: RNase free water
<i>cDNA Sequencing</i>	>150 ng polyA+	>10	>15	260/280 = 2.0 260/230 = 2.0-2.2	-	200 ng cDNA can be used as input DNase digested, buffer: RNase free water
<i>cDNA PCR Sequencing</i>	>10 ng polyA+ or >75 ng Total-RNA	>10	>1	260/280 = 2.0 260/230 = 2.0-2.2	-	DNase digested, buffer: RNase free water
<i>Whole Genome Sequencing</i>						
<i>Ligation Sequencing</i>	>2 μ g	10-50	>40	260/280 = 1.8 260/230 = 2.0-2.2	>30 kb	homogenous HMW DNA, RNase digested, buffer: EB
<i>Native Barcoding</i>	>1 μ g per sample	10-50	>40	260/280 = 1.8 260/230 = 2.0-2.2	>30 kb	homogenous HMW DNA, RNase digested, buffer: EB
<i>Ultra Long Reads</i>	40 μ g	750	>55	260/280 = 1.8 260/230 = 2.0-2.2	>100 kb	homogenous UHMW DNA, RNase digested, buffer: EEB (please inquire about EEB buffer)
<i>Genome Rapid</i>	>400 ng	10	>55	260/280 = 1.8 260/230 = 2.0-2.2	>30 kb	homogenous HMW DNA, RNase digested, buffer: EB
<i>PCR Sequencing</i>	>100 ng	>10 - 50	>2	260/280 = 1.8 260/230 = 2.0-2.2	-	clean, target-specific, buffer: EB

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Oxford Nanopore Long-Read Sequencing						
<i>Amplicon Sequencing</i>						
<i>Amplicons by Ligation</i>	>1 μg per sample	10-50	>40	260/280 = 1.8 260/230 = 2.0-2.2	-	clean, target-specific, buffer: EB
16S	>10 ng	>10	>1	260/280 = 1.8 260/230 = 2.0-2.2	-	clean, target-specific, buffer: EB

Sanger Sequencing

<i>Full Service</i>						
<i>Plasmid</i>	>500 ng	-	100 – 250	260/280 = 1.8 260/230 = 2.0-2.2	<5 kb	Buffer: 10 mM Tris/HCl or water Primer concentration 10 μM
	>800 ng	-	150 – 600	260/280 = 1.8 260/230 = 2.0-2.2	5 kb – 15 kb	Buffer: 10 mM Tris/HCl or water Primer concentration 10 μM
	>1,5 μg	-	>600	260/280 = 1.8 260/230 = 2.0-2.2	>15 kb	Buffer: 10 mM Tris/HCl or water Primer concentration 10 μM
<i>Full Service / Xpress Service</i>						
<i>PCR-product</i>	max. 10 ng	-	<2,5	260/280 = 1.8 260/230 = 2.0-2.2	<100 bp	purified PCR products Primer concentration 10 μM
	50 ng	-	< 5ng	260/280 = 1.8 260/230 = 2.0-2.2	100 bp – 1 kb	purified PCR products Primer concentration 10 μM
	400 ng	-	20 - 50	260/280 = 1.8 260/230 = 2.0-2.2	>1 kb	purified PCR products Primer concentration 10 μM

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Sanger Sequencing						
<i>Xpert Service</i>	-	-	-	-	-	Sample requirements as for Full Service
<i>Pre-mixed Service</i>						
	<i>Plasmid</i>	300 – 600 ng	-	-	-	<10 kb 5 pmol Primer/reaction 7,5 μL total volume
		>700 ng	-	-	-	>10 kb 5 pmol Primer/reaction 7,5 μL total volume
	<i>PCR-products</i>	<100 ng	-	-	-	100 bp – 1 kb 5 pmol Primer/reaction 7,5 μL total volume
		>100 ng	-	-	-	>1 kb 5 pmol Primer/reaction 7,5 μL total volume
<i>Ready-to-load Service</i>						
	<i>Plasmid, PCR-products</i>	-	20	-	-	Purified sequencing reaction

Fragment Analysis

	<i>Ready-to-run</i>	-	20	-	-	<600 bp Fully prepared samples dissolved in formamide
	<i>Pre-pared</i>	-	10	-	-	<600 bp samples dissolved in formamide without length standard
<i>STR analysis</i>		50 ng	-	5	260/280 = 1.8 260/230 = 2.0-2.2	- Buffer: 10 mM Tris/HCl, low TE or water